

Check videos of protocol, examples of results and much more on: idylle-labs.com/brighter

A protocol designed in February 2023.

BrightER PROTOCOL



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1. The material you need

REAGENTS

BrightER 5 mM in DMSO/PEG solution

CONSUMABLES

- Coverslips or imaging chambers (depending on your experiment)
- Cell culture medium

LABWARE

Water-bath heating at 70°C

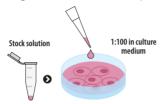
2. Storing the BrightER stock solution

- BrightER can be stored at 4°C for up to one month. For a long-term storage, aliquot and store at -20°C.
- Always keep the vial protected from light.

3. Using BrightER

- Seed your cells on coverslips or imaging chambers and culture as desired.
- We recommend imaging cells at a low density for an optimal visualization of the Endoplasmic Reticulum structure.

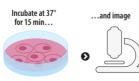
- Take your BrightER stock solution out 20-30 minutes before use to bring it to room temperature.
- Wash cells with PBS to get rid of cellular debris.
- · Add pre-warmed culture medium.
- · Vortex your BrightER stock solution.
- Add the BrightER stock solution, respecting a 1:100 dilution factor in your cell culture medium. The BrightER final concentration is 50µM.



- This concentration should guarantee a bright signal but might need to be adjusted depending on the cell type used.
- Stir manually the dish with circular movements so that the BrightER is dispersed evenly on the cell layer.



Incubate at 37°C for 15 min and proceed immediately to imaging.



- The BrightER fluorescence signal is observable for at least 3 hours.
 If the fluorescence signal is fading, the cell culture medium can be supplemented with additional BrightER.

4. Trouble shooting

- Aggregates may form in the BrightER solution upon storage.
 If necessary, vortex thoroughly for 30 seconds and heat at 70°C for 90 seconds. Doing this twice should help dissolving aggregates.
- You may transfer the BrightER stock solution into an Eppendorf vial to facilitate the heating process when using a water-bath.

