For Research Use Only Not for Diagnostic Use

EasyXpress Viral Nucleic Acid Release Reagent
VB-1000

Introduction

The EasyXpress Viral Nucleic Release reagent is designed for the rapid isolation of small quantities of viral RNA or DNA from cell culture supernatants and biological samples in microfuge tubes or in a 96-well format for subsequent quantitative and real-time RT-PCR analysis. The kit contains VNAR Protease, a proprietary mixture of buffers and proteases that allow for the isolation of up to 96 viral nucleic acid samples in under an hour with no sample filtration or precipitation steps.

Kit Contents

The EasyXpress Viral Nucleic Acid Release Kit contains 10 vials, 1ml each of VNAR protease (1000 samples).

Storage Conditions

VNAR protease should be kept at -20°C.

Product Use Limitations

The EasyXpress Viral Nucleic Acid Release Kit is intended for Research Purposes only, not for Diagnostic Use and we make no warranties or claims whatsoever with regards to the expressed or implied suitability, durability, fitness or utility of this product. In no event shall we be liable for claims for any damages, whether direct, incidental, foreseeable, consequential or special including, but not limited to loss of profit whether based on warranty, contract, in tort, through negligence or in any other manner arising from the use of this product.

Reagents and Equipment to be Supplied by User

Pipetman with tips
Reagent reservoirs for multichannel pipets
Disposable gloves
Table top centrifuge with 96-well plate centrifuge rotor adaptor or microcentrifuge
RNase-and DNase-free water
96-well thermocycler or 37°C and 95°C heating block/water bath.
RNase- and DNase-free microfuge tubes or 96-well plates with cap sealing strips.

Rapid Viral Nucleic Acid Isolation Kit Protocol

1. Add 10 µl of sample of interest per tube/well followed by 10 µl of thawed VNIK Protease.
2. Spin down the samples in the centrifuge (14,000 rpm (16,000g) for 10 sec in a microfuge or 1000g for 1 min in a centrifuge containing a 96-well plate adaptor) .
3. Heat samples to 37°C for 20 min.
4. Heat samples to 94-95°C for 10 min.
5. Place samples on ice for 2 min.
6. Spin down the samples.
7. Add 80 µl RNase- and DNase-free water to the samples and pipet up and down three times to mix. The nucleic acid content of the samples is now 1/10th of that of the original samples. The sample can now be used directly for quantitative and real-time PCR and RT-PCR analysis.[1]

[1] in rare instances, inhibitors of the RT or PCR reaction may remain upon dilution. Inhibiton is indicated by an increase in the total PCR product calculated when the mixture is diluted further- for example in 100X vs. 10X samples. In those instances a further dilution of the mixture beyond the suggested 10X dilution would be required for sample processing.

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